

Curriculum Vitae et Studiorum

PERSONAL INFORMATION

First name: Luigi *Last name:* Leone

Date of Birth: December 22nd 1979 *Place of Birth:* Naples

Street Address: via Miliscola – 80078, Pozzuoli (NA) – Italy

Nationality: Italian

Contacts: mobile +39 3388238212; lab +39 095 7338327; home +39 081 8662570

email: luigi.leone@icb.cnr.it

EDUCATION AND TRAINING

High school: from 1993 to 1999 Qualified Laboratory technician at the “Istituto Tecnico Industriale Statale (ITIS)” of Pozzuoli (NA) Italy – grade 60/100.

WORK EXPERIENCE

October 2002 – October 2003 Technician in the Microbiology Laboratory of the “Istituto di Chimica Biologica (ICB) – (CNR)”, Pozzuoli (NA) – Italy, directed by Dott.ssa Barbara Nicolaus. *Thematic Project:* “Microbiology of Extremophiles and studies on their metabolites.”

January 2004 – April 2009 Project contract as a technician in the Neurobiology of Development Laboratory of the “Istituto di Genetica e Biofisica A. Buzzati Traverso (IGB-ABT) – National Research Council (CNR)”, Naples – Italy, directed by Dott. U. di Porzio.

Thematic Project: “Trophic signalling by GDNF ligands and their receptors in neuronal development and repair”.

May 2009 – August 2010 Project contract as a technician in the facility core of the “Istituto di Genetica e Biofisica A. Buzzati Traverso (IGB-ABT) – National Research Council (CNR)”, Naples – Italy, directed by Dott. A. Baldini.

Thematic Project: Facility Deep sequencing

September 2010 – April 2012 Permanent contract “C.T.E.R. VI level” at “Istituto di Chimica Biomolecolare (ICB) – National Research Council (CNR)”, Catania – Italy

May 2012 – May 2013 Permanent contract “C.T.E.R. VI level” at “Department of Molecular and Human Genetics, Baylor College of Medicine/ Jan and Dan Duncan Neurological Research Institute” – Houston – TX – USA, directed by Dott. Juan Botas

June 2013 – present Permanent contract “C.T.E.R. VI level” at “Istituto di Chimica Biomolecolare (ICB) – National Research Council (CNR)”, Pozzuoli Naples – Italy

RELEVANT SCIENTIFIC TECHNIQUES AND SKILLS ACQUIRED

- Genome analysis

Plasmid & genomic DNA manipulation Cloning

Purification and isolation of genomic DNA from mammalian cells and tissues DNA sequencing

In vitro amplification by PCR In vitro

synthesis of cDNA

Preparation of radiolabel DNA probes

Preparation of samples for hybridization on “DNA chips” Southern Blot and

Dot Blot

Design and test of oligos Luciferase

assays

Techniques of chromatin immunoprecipitation (chIP)

- Gene expression analysis

Total and Messenger RNA

Extraction from cells and tissues

Semiquantitative RT-PCR

Purification and recovery of RNA

Preparation and sequencing of DNA/RNA/mRNA with platform Solid 3plus of Applied Biosystem

- **Nucleic acid electrophoresis**
RNA and DNA agarose gel
- **Functional, structural and expression analyses of proteins in vivo and in vitro**
Biochemical & immunofluorescences analysis SDS-PAGE and Western Blot
Immunofluorescence analysis in eukaryotic cells and tissue by confocal microscopy Handling of expression vectors
Elisa
Quantitative Real-time PCR
- **Handling of prokaryotic cellular system**
Microbiological and biochemical techniques for the cultivation of bacteria
- **Handling of eukaryotic cellular system**
Eukaryotic cell culture & transfection
 - Tissues culture techniques
 - Transient transfection
 - Transfection: Calcium phosphate, Fugene, lipofectamine, electroporation
- **Immunostaining**
Handling of microtome and cryostate instruments Optical and immunofluorescence microscopy Staining on tissue sections and on cell culture
- **Handling and use of animal model**
Mice care and inbred
Stereotactic and intraperitoneal injections Behavioral tests
Heart perfusion
Tissues harvesting and dissection for RNA, DNA and protein extraction
Use of laboratory instruments (quantitative imaging “Typhoon 9400”, spectrophotometer, centrifuges, sterile laminar flow hood, horizontal and vertical, and microscopes)
- **Microbial Culturing Techniques**
Medium preparation, cultures growth and maintenance, axenic cultures preparation, microbial fermentation by using waste biomass as sole carbon source, recovery and characterization of biomolecules (enzymes, lipids and exopolysaccharides)
- **Chromatography Techniques**
Thin layer chromatography (T.L.C.), Fast protein liquid chromatography (F.P.L.C.), High Performance Anion-Exchange. Chromatography with Pulsed. Amperometric Detection (HPAE-PAD).
- **Enzymatic assay:** Determination of reducing sugars (DNS methods) for carbohydrase activities.
- **Good knowledge of software and on-line resources for Bio-informatic analysis**
Excellent knowledge of operating system MacOs and Microsoft Windows Excellent knowledge of Microsoft office
Good knowledge of Factura 2.0, AutoAssembler 2.1, DNA Star, Oligo 4.0 e 6.7, Edit View, Blast/Blat, DNA Strider, Ensemble, Gel Doc di QuantityOne, ImageJ, OMIM, Genome Browser, Unigene, PubMed

Good knowledge of English